

General and species-specific recommendations for minimal requirements for the use of cephalopods in scientific research

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Abstract

Here we enlist species-specific recommendations for housing, care and management of cephalopod molluscs employed for research purposes with the aim of contributing to the standardization of minimum requirements for establishments, care and accommodation of these animals in compliance with the principles stated in Directive 2010/63/EU. Maximizing their psychophysical welfare was our priority. General recommendations on water surface area, water depth and tank shape here reported, represent the outcome of the combined action of the analysis of the available literature and an expertise-based consensus reached – under the aegis of the COST Action FA1301 - among researchers working with the most commonly used cephalopod species in Europe. Information on water supply and quality, environmental conditions, stocking density, feeding and handling are also provided. Through this work we wish to set the stage for a more fertile ground of evidence-based approaches on cephalopod laboratory maintenance, thus facilitating standardization and replicability of research outcomes across laboratories, at the same time maximizing the welfare of these animals.

Introduction

Since January 1st 2013 the use of any live individual (from hatching) belonging to cephalopod molluscs is regulated for scientific purposes in Member States of the European Union by the Directive 2010/63/EU¹ and the national transposed legislations (see Art. 1 3(b)). Cephalopods are the sole invertebrates to have reached the same legal protection analogous to any other vertebrate species, as laboratory animals.²⁻⁴ Their inclusion represents a remarkable turning-point of the current policies, while offering a unique occasion for putting greater attention towards standardization of procedures for the daily care and management of animal welfare as applied to these organisms.^{4,5}

In the current version of the Directive 2010/63/EU and more precisely in part B of the Annex III (Requirements for establishments and for the care and accommodation of animals),¹ a list of mandatory minima for cage sizes, environmental enrichment and group housing - just to mention a few - is available for a number of commonly utilized laboratory species, namely for mice, rats, gerbils, hamsters, guinea pigs, rabbits, cats, dogs, ferrets, non-human primates, farm animals, birds, amphibians, reptiles, and - to a lesser extent - for fishes.¹ Such standards, based on scientific evidence, reflect current practice in Member States of the EU - as a result of the transposed European legislation. It is interesting to note that these are also widely utilized abroad.

When considering fishes, Annex III provides only general information for animals belonging to this taxon (e.g., water supply and quality, lighting, feeding and handling). This contrasts with the previously listed organisms, and the generality of recommendations considering that more than 34,000 different fish species are currently known, and that different species have different physiological and behavioural 'characteristics' (see e.g., FISHBASE: www.fishbase.us/home.htm).⁶

In the case of cephalopods, requirements for the care and accommodation of animals are not available yet.

Cephalopods are characterized by some remarkable features of their Bauplan, physiology and biology.⁷⁻¹¹ They inhabit different geographical areas and diverse environments around the seas of the world. These molluscs account for a wide and diverse variety of taxa also reflecting their adaptations to different habitats in the oceans (e.g., benthic and pelagic, intertidal areas and deep sea, polar regions and the tropics) and at different depth strata.¹¹⁻¹⁶ In addition to the high degree of interspecific variation, many species undergo large physical, physiological, and behavioural changes as a natural part of their life cycle.¹⁷⁻²¹

Their active predatory behaviour include different feeding strategies;¹⁵ these are subjected to repeated changes during behavioural development after hatching for some species, and - relevant to this work - following adaption to captive environment. Few cases of scavenging and filter-feeding habits are also

reported.^{15,22,23} Mating and reproductive strategies cover all possibilities,²⁴ spanning from the high competition for fertilizing females to the random encounter between solitary individuals in the dark deep ocean. Different spawning cycles and strategies have been observed,²⁴ and for some species maternal care has been reported.^{25,26}

To partially support a description of the biological and ecological variety of forms cephalopods represent, Table 1 includes a summary of the different adaptations characterizing the species considered in this work.

Here we provide a set of minimal requirements and recommendations for the care and welfare of cephalopods utilized in scientific research, in compliance with the principles stated in the Directive 2010/63/EU. Information given herein is considered essential for housing and care of the most commonly utilized cephalopod species in EU countries and serves to promote standardization and guidance. This work refers to the 'Guidelines for the treatment of animals in behavioural research and teaching'²⁷ and should be considered complementary to the 'Guidelines for the Care and Welfare of Cephalopods in Research', hereafter referred to as the 'Guidelines'.²⁸

Our goal is to achieve standardisation of minimum requirements for housing, care and accommodation of each cephalopod species of interest. However, this is

challenged by some degree of intraspecific variation and large physiological changes occurring as part of their life cycle (in some species more marked than others), so that recommendations of housing, care and even of applicability of some experimental procedures should consider eventually how to cope at some extent with this variability.

Towards species-specific recommendations for care of cephalopods in research: consensus-based approach and *modus operandi*

Despite the long standing tradition of research on cephalopods, and availability of published accounts about care in controlled conditions, accumulating from at least the early twentieth century,⁵ the absence of species-specific welfare-centred guidelines for the care of these animals led to the adoption of various 'approaches' for their housing, maintenance and care, thus making the level of standardization available for vertebrates²⁹ still far from being achieved for studies involving live cephalopods (for review see De Sio et al.⁵).

Despite the level of accuracy reached in the 'Guidelines' species-specific information for care and management are not included in the appendix 2 (Housing, Environmental Parameters, Transport and Feeding) of Fiorito et al.²⁸

To fill such a gap the [COST Action FA1301](#) and [CephRes](#) promoted and facilitated a consensus-based initiative with the aim of producing a tabularized

recommendation for cephalopods housing, in line with the required content of the Annex III of the Directive 2010/63/EU. This activity has been spanned in a series of meetings and working groups and was initiated in a coordinated way during [CephsInAction meeting in Berlin](#) (Germany: 2016). There, experts convened to discuss about *i.* limitations of the current approaches available in different laboratories for housing and care of cephalopods for scientific research, *ii.* recognized the information included in the Guidelines, *iii.* discussed about the requirements and specific information included in Annex III of the Directive for other species thus to explore a possible template as working method. During the meeting experts began to review published data and white papers in order to find relevant information with the aim to compile a set of cephalopod-care oriented data based on scientific evidence. Information was considered in the sake of the evaluation of the environmental, biological and behavioural needs of different cephalopod species thus to increase success of caring, maintenance, growth and rearing aimed to reduce stress and increase welfare in research establishments. This approach was followed by several remote meetings and exchanges between a selected number of people representing the whole group that, after more than a year, were able to prepare a collation of available information (see below) as a ground for further discussion and refinement; the basis of this compilation are included Appendices A (Environment), B (Accommodation and Care; Enrichment) and C (Thermal, oxygen and salinity tolerance ranges) included in [Supplementary Info](#) in this work.

Coordinated by the COST Action FA1301, the experts met again to achieve consensus and finalize details of the planned work. The occasion was given by one of the last meetings of the COST Action FA1301 ([Galway, Ireland: 2017](#)) also with the aim of monitoring the impact of the transposition of Directive 2010/63/EU and MFSD in EU Member States, COST countries and abroad, facilitate and increase the improvement of the available knowledge on cephalopod biology, physiology and behavioural plasticity that may affect animals' welfare, and contribute to the development of knowledge on care, rearing, environmental and requirements of different cephalopod species to facilitate the standardization in animals' care and increase their welfare. The final step was to refine and discuss some relevant aspects with the Commission during a [CephsInAction Technical Workshop](#) held in Brussels (Belgium: 2017).

Overarching aim of this long work was thus to provide an updated *supplementum* to the 'Guidelines'²⁸ providing more detailed species-specific source of information related to cephalopod care; this resulted to be the basis for the identification of mandatory minima for the housing and care of cephalopods for scientific purposes.

Target Species

The biggest challenge when dealing with cephalopods is the lack of captive breeders cultured for this purpose due to several bottlenecks³⁰ and the consequent need for procuring the specimens from the wild following, sufficient justification, as regulated by the Competent Authority (see Article 9.2¹). Capture and transport of live cephalopods pose many concerns for the welfare of these animals and thus impose undisputable restrictions on the number of species that can be employed in research, at least until standardised and solid protocol will be implemented. The urge for defining the most suitable requirements for cephalopods derives from their particular vulnerability when kept in controlled conditions and their special ecologically-demanding attentions needed for their welfare when housed in a limited space with artificial conditions. As mentioned above, cephalopods are stenohaline and stenotherm thus the range of water quality parameters and environmental settings to which they can be subjected to is very narrow and could easily expose animals to distress and potential suffering if not properly monitored. For such reasons and because animals are considered to have an intrinsic value, EU set strict and clear rules for conducting experiments with laboratory animals, starting from the mandatory requirement of competence of the personnel carrying out procedures (Art. 23¹).

Bearing all this in mind we focus here on the following list of species of interest (see also Tables 1 and 2) for research in EU countries: *Nautilus pompilius*, *N. macromphalus*, *Sepia officinalis*, *S. pharaonis*, *Euprymna scolopes*, *E. berryi*, *E.*

hillebergi, *E. tasmanica*, *Loligo vulgaris*, *L. forbesii*, *Sepioteuthis sepioidea*, *S. lessoniana*,
Octopus vulgaris, *O. bimaculoides*, *O. maya*. The species considered in this list do
not exclude other representatives, congeneric and/or belonging to the same taxa,
characterized by similar life styles and adaptations. We are committed to expand
lists and biological information included in the tabularized overviews (e.g. Table
1 and Supplementary Appendix C) based on scientific evidence.

Source of data and extraction of information

Information about care and housing parameters adopted for cephalopods that
we consider herein derives from an expert analysis of various sources (a total of
97 different journal papers, reviews and book chapters) and a few *ad hoc* studies
reporting details considered robust enough to justify their inclusion in our
analysis. As mentioned in [Supplementary Info](#) to this work, we have also
taken into account the classic contribution made by Grimpe,³¹ which
represents the first available set of guidelines for laboratory rearing and
maintenance of cephalopods intended to be studied in zoology and
physiology,⁵ together with other reviews on care of cephalopods.^{28,32-39}

Information and other data collected were collated in a tabularized overview of
the ‘care and accommodation of animals’, providing important details regarding
environment (water supply and quality, lighting, noise and vibration),

accommodation, care (housing, stocking density and sex ratio, diet), enrichment, and temperature, oxygen and salinity tolerance ranges (see Appendices A-C in [Supplementary Info](#), respectively). All the information collected was accurately curated in order to be considered of relevance for the aims of this work and for the ultimate goal of producing reference to recommendations for the care and welfare of the target species.

Based on the above collection and through the joint effort of researchers providing different expertise, we produced a consensus-based set of standardized values and parameters required to accommodate the selected cephalopod species here considered. Data were revised by experts working with each selected species and then agreed upon during scheduled designated meetings. Whenever species-specific information was missing, we completed it with proper indications based on the working experience of the experts and the current good practices for each specific cephalopod taxon employed in research. The recommended notes here included are based on accurate data mining from our source of information (see Appendices A-C in [Supplementary Info](#)) and discussion between experts at dedicated meetings. The above-mentioned appendices represent background information on the species-specific proposal of recommendations for care of cephalopods for scientific purposes.

The following notes are inspired by the criteria adopted for Urodeles and Anurans amphibians as included in Annex III of the Directive. The approach we

followed is similar to that included in the “Background information on the species-specific proposals for reptiles presented by the Expert Group on Amphibians and Reptiles”⁴⁰ provided to the Council of Europe in order to implement the species-specific section about amphibians and reptiles in the Annex III of the Directive 2010/63/EU.¹

Requirements for housing and care of live cephalopods

Table 2 lists the recommended minimum requirements for housing live cephalopods under research settings. Values included are based on our data-source (see Appendices A-C in [Supplementary Info](#)), and other relevant publications, including considerations about health monitoring and post-mortem evaluation of cephalopods.³⁹ It is interesting to mention here that hatchlings viability, appropriate water quality and food items provided to cephalopods, and suitable living space were considered essential requirements by Grimpe³¹ and years afterwards by von Boletzky³⁸ for the aim of achieving the best possible conditions for the care and management of live cephalopods for scientific research purposes.

229 General Recommendations

230 Part A - General section - of Annex III of the Directive 2010/63/EU includes
231 general recommendations about the overall care and accommodation for
232 laboratory animals. Such guidance highlights that the welfare of the reared
233 species depends first of all on facility and holding rooms status together with the
234 environment and general care to which all subjects are housed, and finally on the
235 individual husbandry conditions (Part A, Annex III¹).

236 When referring to cephalopod molluscs, it is easy to understand that a crucial
237 element to consider is the system that supplies seawater of adequate quality. In
238 general, either open or closed systems are adopted. Each one has its pros and
239 cons and the choice of one over the other should be properly and attentively
240 made (see [Supplementary Info](#)). Water temperature should be set and regulated
241 according to the natural range for the species and the geographical area in which
242 it was captured (if from the wild); the life-stage should also be considered.

243

244 Regardless of the cephalopod species, a daily monitoring of the welfare state
245 should be carried out by competent personnel in appropriate rooms where
246 «simple diagnostic tests, post-mortem examinations, and/or the collection of
247 samples can be performed» (see 1.3(a) of Part A, Annex III¹); «these checks shall
248 ensure that all sick or injured animals are identified and appropriate action is
249 taken» (*ibid.*, see 3.1(b)).

Noise and other interference should always be kept to a minimum to avoid or reduce any cause of potential distress in animals.²⁸ The lighting conditions should be regulated as well, to respect and emulate the biological and physiological needs of cephalopods.

As for animals housing, concerns should be made towards the species-specific “social” needs (i.e. group- or solitary-living habits of the species) and also their behavioural requirements, providing access to adequate environmental enrichment that fosters visual, tactile and cognitive stimulation and at the same time attempts to reproduce the motivation standards to which the animal is exposed to in nature. Resting and ‘sleeping’ areas should be a minimum requirement to help specimens feel protected while inactive, together with presenting dens in which to hide, or sandy areas to dig themselves into whenever they feel threatened by potential external menaces.

Concerning feeding it is required that animals should have access to a diet that meets their nutritional and behavioural needs in form, content and presentation. The food shall be palatable and non-contaminated with chemical, physical and microbiological traces (see 3.4(a, b)¹). General principles about the type of food items to provide to animals is included in Fiorito and coworkers.²⁸

Each animal shall be able to access food, allowing to exhibit the species-specific predatory behaviour and sufficient feeding space to limit competition and

‘cannibalism’. We recommend that *ad libitum* feeding should be avoided to prevent motivational decline and reduce risks of counter effects in the efficiency of the systems assuring adequate quality of seawater.

Species-Specific Requirements

As mentioned above, there are no cephalopod species-specific requirements included in part B of Annex III of the Directive 2010/63/EU. Also, information provided for fish are very general and not adequate to translate to cephalopods. Here we include (Table 2) a ‘minimum water surface area’ per animal for each of the species considered. This has been defined as «a constant function of the “footprint” area of the animal, at least within a given species». The animal footprint area refers to the square of any linear dimension, creating a plot of the amount of space allocated to animals, able, on a proportional basis, to take into account both large and small subjects.⁴⁰ In addition, this minimum space requirement allows for the introduction of environmental enrichment for a given specimen. When the species in question can be group-held, a value for the minimum water surface area for each additional animal has to be defined, taking into account the biological tolerance in terms of inter-individual space requirements of the target species.

In identifying tank shape and the minimum water depth we also considered the species’ physiological and behavioural needs such as their benthic or pelagic

biology or the daily vertical migration (with the unavoidable constraint of captivity).

More in detail, parameters included in Table 2 referring to a specific taxon are proposed taking into account different body size (as dorsal mantle length for a coleoid, and as shell diameter for Nautilidae). Furthermore, the minimum water surface area refers generally to life stages that do not consider the transition from the paralarval form as it may require additional considerations⁴¹ and therefore any value should be readapted according to a scaling principle.

As for the tank, any shape can be chosen with few exceptions that may concern animals' lifestyle. As can be seen from the "minimum water depth" reported in Table 2, Nautilidae need a tank that is consistently much higher than wider when compared to coleoids as they usually perform daily vertical migrations.⁴² Benthic species usually require large ground areas, low water column and rounded edges, while pelagic species need circular (or oval, as for squid) tanks, smaller areas, with sufficient space to swim or high-water column, thus allowing the animal to display its natural behaviour. To this regard, families such as Octopodidae need dens and refuges as minimum requirement, in order to make the animals free to express their natural behaviour, including hiding in a den. Other cephalopods, and in particular female specimens require bottom substrate as a spawning ground or surfaces suitable for attaching egg masses. In addition, environmental enrichment and feeding protocols best fitting the behaviour of

these animals should be provided in order to reproduce as much as possible their natural environments while keeping their motivation high (see General Recommendations above).

Little information is currently available on the minimum water surface area for single and multiple specimens and the proposed values derive from the critical analysis of data available in the scientific literature. In some instances, information derived from studies carried out in other contexts (i.e. aquaculture or public display) where the maximising the growth rate, and the stocking density or longevity are the main focus. As a consequence, we discussed about the available information and attempted to best fit cephalopods' biology requirements and maximise the achievement of the Five Freedoms.^{43,44}

The number of specimens that can be hold in the same tank depends on species-specific social behaviour, age and size of the animals and individual space requirements in terms of water surface area. For instance, *O. vulgaris* is generally a solitary-living species⁴⁵ and therefore should be kept isolated and, if group housing is inevitable, visual interaction must be avoided or maximally reduced with at least a shelter per individual together with larger space as indicated by the parameters under the entry "minimum water surface area for each additional animal in group-holding". Nevertheless, the lack of scientific data makes it difficult to define categoric recommendations which are therefore based on prolonged practical experience in successful rearing of cephalopods.

Closing Remarks

Cephalopods, though accounting for less than 2% of the phylum of Mollusca, are considered as the class of invertebrates endowed with the largest nervous system underlying levels of plasticity comparable to those of some vertebrate. Their sentience (*sensu* Broom⁴⁶) is at the basis of the decision of including them as the sole invertebrate taxon listed in the Directive 2010/63/EU. A recent framework for evaluating scientific evidence of sentience based on eight criteria provided further support.⁴⁷

Very little is known about inter-specific compatibility, that appear to be a possible scenario in public display, when multiple species are housed in the same enclosures. Studies are required with focus on animal welfare, and these can be facilitated by works carried out in the wild, but also when accurate welfare monitoring are applied to public aquaria.

For the first time, we attempted at providing species-specific minimum requirements for housing, accommodation, and care of live cephalopods with the aim to support inclusion of required information in Annex III of the Directive. This work represents a step forward towards the improvement of cephalopods welfare. Recommendations proposed are intended to enhance the current available knowledge on this taxon biology, husbandry and care to guide both experienced and early career scientists, students, caretakers, technicians and veterinarians towards the achievement of best practices for increasing the success

in cephalopods' maintenance, growth and rearing for scientific purpose in research establishments.

Requirements listed in Table 2 derived by a joint effort of a group of experts - based on the supporting information (see Appendices A-C in [Supplementary Info](#)) - coordinated for matching and adapting the available data on space requirements with the experience and knowledge on the behavioural and physiological needs of cephalopod, keeping animal welfare as our priority. The consensus-based approach here adopted represents one of the main outcomes of the COST Action FA1301 and originates from the whole series of meetings starting in late 2014.

Here we reviewed all the available literature and collected relevant data about the use of cephalopods as experimental animals. We also provided supplementary information regarding water quality requirements, feeding and environmental needs of the selected species in order to guide researchers and caretakers in fully achieving animals' welfare while attempting to implement the Five Freedoms principles to cephalopods.

The guidance list of minimal requirements for housing and care of selected cephalopod species included in this work has to be considered as complementary to the general Guidelines²⁸. It represents a further step towards a more intense series of studies providing adequate, experimentally-based data helping to refine the requirements here included.

Although might be demanding for researchers and establishments, the minimum accommodation needs represent the basis to ensure animal welfare and consequently improve the quality of research data by making it more transparent, replicable and thus reliable.

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Data Availability

This work is based on previously published works and extensively refer to those. No primary data have been utilized in this review. [Supplementary material](#) includes detailed information and references for original data.

Reference List

1. European P and Council of the European U. Directive 2010/63/EU of the European Parliament and of the Council of 22 September 2010 on the protection of animals used for scientific purposes, (2010, <https://eur-lex.europa.eu/legal-content/EN/ALL/?uri=CELEX:32010L0063>).
2. Smith JA, Andrews PL, Hawkins P, et al. Cephalopod research and EU Directive 2010/63/EU: Requirements, impacts and ethical review. *Journal of Experimental Marine Biology and Ecology* 2013; 447: 31-45.
3. Di Cristina G, Andrews P, Ponte G, et al. The impact of Directive 2010/63/EU on cephalopod research. *Invertebrate Neuroscience* 2015; 15: 8.
4. Ponte G, Andrews P, Galligioni V, et al. Cephalopod Welfare, Biological and Regulatory Aspects: An EU Experience. In: Carere C and Mather J (eds) *The Welfare of Invertebrate Animals*. Cham, Switzerland: Springer International Publishing, 2019, pp.209-228.
5. De Sio F, Hanke FD, Warnke K, et al. E Pluribus Octo–Building Consensus on Standards of Care and Experimentation in Cephalopod Research; a Historical Outlook. *Frontiers in physiology* 2020; 11: 645.
6. Klimpel S, Kuhn T, Münster J, et al. Anatomy and Morphology of Fish and Cephalopods. In: Klimpel S, Kuhn T, Münster J, et al. (eds) *Parasites of Marine Fish and Cephalopods: A Practical Guide*. Cham: Springer International Publishing, 2019, pp.15-27.
7. Hanlon R, Vecchione M and Allcock L. *Octopus, Squid, and Cuttlefish: A Visual, Scientific Guide to the Oceans' Most Advanced Invertebrates*. University of Chicago Press, 2018.
8. Shigeno S, Takenori S and Boletzky Sv. The origins of cephalopod body plans: a geometrical and developmental basis for the evolution of vertebrate-like organ systems. In: Tanabe K, Shigeta Y, Sasaki T, et al. (eds) *Cephalopods - Present and Past*. Tokyo: Tokai University Press, 2010, pp.23-34.
9. Boletzky Sv. Early ontogeny and evolution: The cephalopod model viewed from the point of developmental morphology. *Geobios* 1989; 22: 67-78. DOI: [https://doi.org/10.1016/S0016-6995\(89\)80008-7](https://doi.org/10.1016/S0016-6995(89)80008-7).
10. Budelmann BU, Schipp R and Boletzky Sv. Cephalopoda. In: Harrison FW and Kohn AJ (eds) *Microscopic Anatomy of Invertebrates*. New York: Wiley-Liss, Inc., 1997, pp.119-414.
11. Packard A. Cephalopods and fish: the limits of convergence. *Biol Rev* 1972; 47: 241-307.

- 439 12. Jereb P, Roper C, Norman M, et al. *Cephalopods of the World. An Annotated and*
440 *Illustrated Catalogue of Species Known to Date. Volume 3. Octopods and Vampire Squids.*
441 Roma, Italy: FAO, Food and Agriculture Organization of the United Nations, 2016,
442 p.370.
- 443 13. Jereb P and Roper C. *Cephalopods of the world. An annotated and illustrated*
444 *catalogue of species known to date. Volume 1. Chambered Nautiluses and Sepioids (Nautilidae,*
445 *Sepiidae, Sepiolidae, Sepiadariidae, Idiosepiidae and Spirulidae).* Rome, Italy: FAO, Food and
446 Agriculture Organization of the United Nations, 2005, p.262.
- 447 14. Jereb P and Roper C. *Cephalopods of the world. An annotated and illustrated*
448 *catalogue of cephalopod species known to date. Volume 2. Myopsid and Oegopsid Squids.*
449 Rome, Italy: FAO, Food and Agriculture Organization of the United Nations, 2010,
450 p.605.
- 451 15. Villanueva R, Perricone V and Fiorito G. Cephalopods as predators: a short
452 journey among behavioral flexibilities, adaptations, and feeding habits. *Frontiers in*
453 *Physiology* 2017; 8: 598.
- 454 16. Ponte G, Taite M, Borrelli L, et al. Cerebrotypes in Cephalopods: Brain
455 Diversity and Its Correlation With Species Habits, Life History, and Physiological
456 Adaptations. *Frontiers in Neuroanatomy* 2020; 14: 105.
- 457 17. Rosa R, Pissarra V, Borges FO, et al. Global Patterns of Species Richness in
458 Coastal Cephalopods. *Frontiers in Marine Science* 2019; 6. Original Research. DOI:
459 10.3389/fmars.2019.00469.
- 460 18. Sweeney MJ, Roper CF, Mangold KM, et al. "Larval" and juvenile cephalopods:
461 a manual for their identification. *Smithson Contrib Zool* 1992: 1-282.
- 462 19. Young RE and Harman RF. "Larva", "paralarva" and "subadult" in cephalopod
463 terminology. *Malacologia* 1988; 29: 201-207.
- 464 20. Boletzky Sv. Biology of Early Life Stages in Cephalopod Molluscs. In:
465 Southward AJ, Tyler PA, Young CM, et al. (eds) *Advances in Marine Biology*. Academic
466 Press, 2003, pp.143-203.
- 467 21. Robin J-P, Roberts M, Zeidberg L, et al. Chapter Four - Transitions During
468 Cephalopod Life History: The Role of Habitat, Environment, Functional Morphology
469 and Behaviour. In: Vidal EAG (ed) *Advances in Marine Biology*. Academic Press, 2014,
470 pp.361-437.
- 471 22. Budelmann BU. Active marine predators: the sensory world of cephalopods.
472 *Marine & Freshwater Behaviour & Phy* 1996; 27: 59-75.
- 473 23. Hanlon RT and Messenger JB. *Cephalopod Behaviour*. 2 ed. Cambridge:
474 Cambridge University Press, 2018.

- 475 24. Rocha F, Guerra Á and González ÁF. A review of reproductive strategies in
476 cephalopods. *Biological Reviews* 2001; 76: 291-304.
- 477 25. Seibel BA, Robison BH and Haddock SHD. Post-spawning egg care by a squid.
478 *Nature* 2005; 438: 929-929. DOI: 10.1038/438929a.
- 479 26. Bush SL, Hoving HJT, Huffard CL, et al. Brooding and sperm storage by the
480 deep-sea squid *Bathyteuthis berryi* (Cephalopoda: Decapodiformes). *Journal of the*
481 *Marine Biological Association of the United Kingdom* 2012; 92: 1629.
- 482 27. Buchanan K, Burt de Perera T, Carere C, et al. Guidelines for the treatment of
483 animals in behavioural research and teaching. *Animal Behaviour* 2021; 171: I-XI. DOI:
484 [https://doi.org/10.1016/S0003-3472\(20\)30373-0](https://doi.org/10.1016/S0003-3472(20)30373-0).
- 485 28. Fiorito G, Affuso A, Basil J, et al. Guidelines for the Care and Welfare of
486 Cephalopods in Research -A consensus based on an initiative by CephRes, FELASA
487 and the Boyd Group. *Laboratory Animals* 2015; 49: 1-90. DOI: 10.1177/0023677215580006.
- 488 29. Hawkins P, Dennison N, Goodman G, et al. Guidance on the severity
489 classification of scientific procedures involving fish: Report of a Working Group
490 appointed by the Norwegian Consensus-Platform for the Replacement, Reduction and
491 Refinement of animal experiments (Norecopa). *Laboratory animals* 2011; 45: 219-224.
492 DOI: 10.1258/la.2011.010181.
- 493 30. Jacquet J, Franks B, Godfrey-Smith P, et al. The Case Against Octopus Farming.
494 *Issues in Science and Technology* 2019; 35: 37-44.
- 495 31. Grimpe G. Pflege, Behandlung und Zucht der Cephalopoden für zoologische
496 und physiologische Zwecke. In: Überhalden E (ed) *Handbuch der biologischen*
497 *Arbeitsmethoden*. Berlin, Wien: Verlag Urban & Schwarzenberg, 1928, pp.331-402.
- 498 32. Boletzky Sv and Hanlon RT. A Review of the Laboratory Maintenance, Rearing
499 and Culture of Cephalopod Molluscs. *Memoirs of the National Museum of Victoria* 1983;
500 44: 147-186.
- 501 33. Boyle PR. *The UFAW handbook on the care and management of cephalopods in the*
502 *laboratory*. Potters Bar Universities Federation for Animal Welfare, 1991, p.915.
- 503 34. Fiorito G, Affuso A, Anderson DB, et al. Cephalopods in neuroscience:
504 Regulations, Research and the 3Rs. *Invertebrate Neuroscience* 2014; 14: 13-36.
- 505 35. Hanlon RT. Maintenance, rearing, and culture of teuthoid and sepioid squids.
506 *Squid as experimental animals*. Springer, 1990, pp.35-62.
- 507 36. Iglesias J, Fuentes L and Villanueva R. *Cephalopod culture*. Springer Science &
508 Business Media, 2014, p.494.

37. Vidal EAG, Villanueva R, Andrade JP, et al. Chapter One - Cephalopod Culture: Current Status of Main Biological Models and Research Priorities. In: Vidal EAG (ed) *Advances in Marine Biology*. Academic Press, 2014, pp.1-98.
38. Boletzky Sv. A brief survey of cephalopod culture techniques. *Turkish Journal of Aquatic Life* 2004; 2: 229-240.
39. Oestmann DJ, Scimeca JM, Forsythe J, et al. Special Considerations for Keeping Cephalopods in Laboratory Facilities. *Journal of the American Association for Laboratory Animal Science* 1997; 36: 89-93.
40. Ewert J-P, Cooper JE, Langton T, et al. *Species specific provisions for Amphibians. Background information for the proposals presented by the Group of Experts on Amphibians and Reptiles*. Report no. GT 123 (2004) 14, 27/08/2004 2004. Strasbourg: Council of Europe.
41. Deryckere A, Styfhals R, Vidal EAG, et al. A practical staging atlas to study embryonic development of *Octopus vulgaris* under controlled laboratory conditions. *BMC Developmental Biology* 2020; 20: 1-18.
42. Barord GJ and Basil JA. *Nautilus. Cephalopod culture*. Springer, 2014, pp.165-174.
43. Webster AJF. Farm Animal Welfare: the Five Freedoms and the Free Market. *The Veterinary Journal* 2001; 161: 229-237. DOI: <https://doi.org/10.1053/tvj.2000.0563>.
44. Mellor DJ. Updating Animal Welfare Thinking: Moving beyond the “Five Freedoms” towards “A Life Worth Living”. *Animals* 2016; 6. DOI: 10.3390/ani6030021.
45. Wells MJ. *Octopus: physiology and behaviour of an advanced invertebrate*. Springer Science & Business Media, 1978.
46. Broom DM. *Sentience and animal welfare*. CABI, 2014.
47. Birch J, Burn C, Schnell A, et al. *Review of the Evidence of Sentience in Cephalopod Molluscs and Decapod Crustaceans*. 2021. London: London School of Economics and Political Science
48. Koestler RJ, Santoro ED and Dingerkus G. Elemental and Ultrastructural Characteristics of the Egg Capsules of *Nautilus pompilius*. *Scanning Electron Microscopy* 1986; 1986: 19.
49. Arnold JM. Reproduction and embryology of *Nautilus*. *Nautilus*. Springer, 2010, pp.353-372.
50. Carlson BA. Collection and aquarium maintenance of *Nautilus*. In: N.H SWB and Landman (eds) *Nautilus*. Springer, 2010, 1991, pp.563-578.

- 542 51. Zhang Y, Mao F, Mu H, et al. The genome of *Nautilus pompilius* illuminates eye
543 evolution and biomineralization. *Nature Ecology & Evolution* 2021. DOI: 10.1038/s41559-
544 021-01448-6.
- 545 52. Willey A. The Oviposition of *Nautilus macromphalus*. *Proceedings of the Royal*
546 *Society of London* 1896; 60: 467-471.
- 547 53. Saunders WB. The species of *Nautilus*. *Nautilus*. Springer, 2010, pp.35-52.
- 548 54. Laptikhovsky V, Salman ALP, Önsoy B, et al. Fecundity of the common
549 cuttlefish, *Sepia officinalis* L. (Cephalopoda, Sepiida): a new look at the old problem.
550 *Scientia Marina* 2003; 67: 279-284.
- 551 55. Villanueva R, Vidal EAG, Fernández-Álvarez FÁ, et al. Early Mode of Life and
552 Hatchling Size in Cephalopod Molluscs: Influence on the Species Distributional
553 Ranges. *PLOS ONE* 2016; 11: e0165334. DOI: 10.1371/journal.pone.0165334.
- 554 56. Messenger JB. The visual attack of the cuttlefish, *Sepia officinalis*. *Animal*
555 *Behaviour* 1968; 16: 342-357.
- 556 57. Nabhitabhata J. *Sepia pharaonis*. In: Iglesias J, Fuentes L and Villanueva R (eds)
557 *Cephalopod culture*. Dordrecht: Springer, 2014, pp.205-224.
- 558 58. Nabhitabhata J and Nilaphat P. Life cycle of cultured pharaoh cuttlefish, *Sepia*
559 *pharaonis* Ehrenberg, 1831. *Phuket Marine Biological Center Special Publication* 1999; 19:
560 25-40.
- 561 59. Lee M-F, Lin C-Y, Chiao C-C, et al. Reproductive behavior and embryonic
562 development of the pharaoh cuttlefish, *Sepia pharaonis* (Cephalopoda: Sepiidae).
563 *Zoological Studies* 2016; 55.
- 564 60. Choe S. On the eggs, rearing, habits of the fry, and growth of some
565 cephalopods. *Bulletin of Marine Science* 1966; 16: 330-347.
- 566 61. Nabhitabhata J and Nishiguchi MK. *Euprymna hyllebergi* and *Euprymna*
567 *tasmanica*. *Cephalopod Culture*. Springer, 2014, pp.253-269.
- 568 62. Nabhitabhata J, Nilaphat P, Promboon P, et al. Life cycle of cultured bobtail
569 squid, *Euprymna hyllebergi* Nateewathana, 1997. *Phuket Mar Biol Cent Res Bull* 2005; 66:
570 351-365.
- 571 63. Lee P, Callaerts P and de Couet H. The Embryonic Development of the
572 Hawaiian Bobtail Squid (*Euprymna scolopes*). *Cold Spring Harbor protocols* 2009; 2009:
573 pdb.ip77. DOI: 10.1101/pdb.ip77.
- 574 64. Singley CT. *Euprymna scolopes*. In: Boyle PR (ed) *Cephalopod Life Cycles*. London:
575 Academic Press, 1984.

65. Hanlon RT, Claes MF, Ashcraft SE, et al. Laboratory culture of the sepiolid squid *Euprymna scolopes*: a model system for bacteria-animal symbiosis. *The Biological Bulletin* 1997; 192: 364-374.
66. Belcaid M, Casaburi G, McAnulty SJ, et al. Symbiotic organs shaped by distinct modes of genome evolution in cephalopods. *Proceedings of the National Academy of Sciences* 2019; 116: 3030-3035. DOI: 10.1073/pnas.1817322116.
67. Norman MD and Lu CC. Redescription of the Southern Dumpling Squid *Euprymna tasmanica* and a Revision of the Genus *Euprymna* (Cephalopoda: Sepiolidae). *Journal of the Marine Biological Association of the United Kingdom* 1997; 77: 1109-1137. 2009/05/11. DOI: 10.1017/S0025315400038662.
68. Squires ZE, Norman MD and Stuart-Fox D. Mating behaviour and general spawning patterns of the southern dumpling squid *Euprymna tasmanica* (Sepiolidae): a laboratory study. *Journal of Molluscan Studies* 2013; 79: 263-269.
69. Porteiro FM, Martins HR and Hanlon RT. Some observations on the behaviour of adult squids, *Loligo forbesi*, in captivity. *Journal of the Marine Biological Association of the United Kingdom* 1990; 70: 459-472.
70. Nabhitabhata J and Ikeda Y. *Sepioteuthis lessoniana*. In: Iglesias J, Fuentes L and Villanueva R (eds) *Cephalopod culture*. Dordrecht: Springer, 2014, pp.315-347.
71. LaRoe ET. The culture and maintenance of the loliginid squids *Sepioteuthis sepioidea* and *Doryteuthis plei*. *Marine Biology* 1971; 9: 9-25.
72. Arnold JM. Observations on the mating behavior of the squid *Sepioteuthis sepioidea*. *Bulletin of Marine Science* 1965; 15: 216-222.
73. Silva L, Sobrino I and Ramos F. Reproductive biology of the common octopus, *Octopus vulgaris* Cuvier, 1797 (Cephalopoda: Octopodidae) in the Gulf of Cádiz (SW Spain). *Bulletin of Marine Science* 2002; 71: 837-850.
74. Avendaño O, Roura Á, Cedillo-Robles CE, et al. *Octopus americanus*: a cryptic species of the *O. vulgaris* species complex redescribed from the Caribbean. *Aquatic Ecology* 2020; 54: 909-925. DOI: 10.1007/s10452-020-09778-6.
75. Zarrella I, Herten K, Maes GE, et al. The survey and reference assisted assembly of the *Octopus vulgaris* genome. *Sci Data* 2019; 6: 13. DOI: 10.1038/s41597-019-0017-6.
76. Forsythe JW and Hanlon RT. Behavior, body patterning and reproductive biology of *Octopus bimaculoides* from California. *Malacologia* 1988; 29: 41-55.
77. Ibarra-García LE, Mazón-Suástegui JM, Rosas C, et al. Morphological and physiological changes of *Octopus bimaculoides*: from embryo to juvenile. *Aquaculture* 2018; 497: 364-372.

- 611 78. Forsythe JW and Hanlon RT. Effect of temperature on laboratory growth,
612 reproduction and life span of *Octopus bimaculoides*. *Marine Biology* 1988; 98: 369-379.
613 DOI: 10.1007/BF00391113.
- 614 79. Albertin CB, Simakov O, Mitros T, et al. The octopus genome and the evolution
615 of cephalopod neural and morphological novelties. *Nature* 2015; 524: 220-224.
- 616 80. Rosas C, Gallardo P, Mascaró M, et al. *Octopus maya*. *Cephalopod culture*.
617 Springer, 2014, pp.383-396.
- 618 81. Avila-Poveda OH, Koueta N, Benítez-Villalobos F, et al. Reproductive traits of
619 *Octopus maya* (Cephalopoda: Octopoda) with implications for fisheries management.
620 *Molluscan Research* 2016; 36: 29-44.
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Table 1. The species considered in this work and their main biological features. The information provided herein serves to depict biological diversity, main physiological and ecological adaptations and some behavioural requirements.

Abbreviations – P: Planktonic paralarvae phase; M: miniature adult (i.e. animals at hatching with most adult characters present in the newly hatched young); Ac: Active predators; S: scavengers; Ad: Adaptable to inert or artificial food; L: Live, selected prey; A: semelparous or simultaneous terminal spawning; B: iteroparous; (*p*) polycyclic spawning; (*m*) multiple spawning; (*in*) intermittent spawning; (*c*) continuous spawning; N/A: Not Available.

Species	Length of egg capsule (mm)	Egg number	Habits of hatchlings ⁱⁱ	Geographical distribution	Depth range (m)	Feeding habits ⁱⁱⁱ	Reproductive strategy ^{iv}	Genome availability
<i>Nautilus pompilius</i>	20.0-40.0 ⁴⁸	10–15 (per year) ^{v, 49}	M ⁴⁹	Indo-West Pacific; Andaman Islands, Ambon, the Philippines, New Guinea to Fiji; north eastern and north western Australia ¹³	0-750 ¹³	S ⁵⁰ , Ad ^{15,36}	B, <i>p</i> ²⁴	Yes ⁵¹
<i>N. macromphalus</i>	~ 45.0 ⁵²	10–15 (per year) ^{v, 49}	M ⁴⁹	Southwestern Pacific Ocean; northeaster Australia, New	0-500 ¹³	S ⁵³ , Ad ¹⁵	B, <i>p</i> ²⁴	N/A

ⁱⁱ See Young & Harman (1988)²¹ for details about the definition of the term paralarvae in cephalopods

ⁱⁱⁱ For further details on the different hunting strategies of adult cephalopods see Villanueva et al. (2017)⁵⁹

^{iv} All the terms are conceived *sensu* Rocha et al. (2001)⁶⁶

^v This information is mainly based on data collected in captivity

Species	Length of egg capsule (mm)	Egg number	Habits of hatchlings ⁱⁱ	Geographical distribution	Depth range (m)	Feeding habits ⁱⁱⁱ	Reproductive strategy ^{iv}	Genome availability
				Caledonia and Loyalty Islands ¹³				
<i>Sepia officinalis</i>	8.0-10.0 ¹³	150-8,000 ^{13,54}	M ⁵⁵	Eastern Atlantic and Mediterranean Sea ¹³	100-200 ¹³	Ac, Ad ^{15,36,56}	B, in ²⁴	N/A
<i>S. pharaonis</i>	24.0-35.0 ⁵⁷	500–3,000 ^{57,58}	M ⁵⁵	Indian Ocean and western Pacific ¹³	10-130 ¹³	Ac, Ad ^{15,36}	B, in ⁵⁹	N/A
<i>Euprymna berryi</i>	3.0-7.7 ⁶⁰	20-300	P ⁵⁵	Indo-Pacific: along coasts of China, south to Hong Kong and Japan, Taiwan, possibly Andaman Islands and Sri Lanka ¹³	Up to 107 ¹³	Ac ⁶⁰	B, m, (in ²⁴)	N/A
<i>E. hyllebergi</i>	3.0-4.0 ⁶¹	100-470 ⁶¹	P ⁵⁵	Eastern Indian Ocean: Thailand, Andaman Sea ¹³	Up to 74 ¹³	Ac, Ad ³⁶	B, c, in ^{24,62}	N/A
<i>E. scolopes</i>	1.5-2.0 ⁶³	12- 300 ⁶⁴	P ⁵⁵	Central Pacific: Hawaiian Islands ¹³	Shallow coastal waters ¹³	Ac ¹⁵	B, c, in ^{24,65}	Yes ⁶⁶
<i>E. tasmanica</i>	3.0-4.0 ⁶¹	100-250 ⁶¹	M ⁶¹	Southern Indo-Pacific: eastern and south eastern Australia ¹³	0-84 ⁶⁷	Ac, Ad ³⁶	B, c, in ^{24,68}	N/A
<i>Loligo vulgaris</i>	2.0-4.0 ¹⁴	1,000-100,000 ¹⁴	P ⁵⁵	Eastern Atlantic Ocean and Mediterranean Sea ¹⁴	100-500 ¹⁴	Ac ¹⁵ , Ad ³⁶ , L	B, in ²⁴	N/A

Species	Length of egg capsule (mm)	Egg number	Habits of hatchlings ⁱⁱ	Geographical distribution	Depth range (m)	Feeding habits ⁱⁱⁱ	Reproductive strategy ^{iv}	Genome availability
<i>L. forbesii</i>	1.8-3.0 ¹⁴	1,000-23,000 ¹⁴	P ⁵⁵	Eastern North Atlantic and Mediterranean Sea ¹⁴	50-1000 ¹⁴	Ac, L ⁶⁹	B, in ²⁴	N/A
<i>Sepioteuthis lessoniana</i>	2.0-3.0 ¹⁴	400–3,500 ⁷⁰	P ⁵⁵	Indo-West Pacific Ocean and Eastern Mediterranean Sea ¹⁴	0-100 ¹⁴	Ac ¹⁵ , Ad ^{36,70}	B, in ²⁴	N/A
<i>S. sepioidea</i>	5.0-6.0 ¹⁴	3-4 per capsule ¹⁴	P ⁵⁵	Tropical western Atlantic Ocean from Cape Canaveral, Florida, Bermuda and the Bahama Islands, Florida Keys, through the Caribbean Islands, Campeche, Yucatan and Venezuela to Farol de Barra Beach, north eastern Brazil ¹⁴	3-20 ¹⁴	Ac ¹⁵ , L ⁷¹	A ⁷²	N/A
<i>Octopus vulgaris</i>	1.4-2.3 ⁷³	100,000-500,000 ¹²	P ⁵⁵	Cosmopolitan; ¹² but see discussion on <i>O. vulgaris</i> species complex ⁷⁴	0-250 ¹²	Ac ¹⁵ , Ad ³⁶	A ²⁴	Yes ⁷⁵

Species	Length of egg capsule (mm)	Egg number	Habits of hatchlings ⁱⁱ	Geographical distribution	Depth range (m)	Feeding habits ⁱⁱⁱ	Reproductive strategy ^{iv}	Genome availability
<i>O. bimaculoides</i>	10.0-17.0 ⁷⁶	~800 ¹²	M ⁵⁵	Northeast Pacific, California (San Simeon) and the California Channel Islands south to at least Guerrero Negro, on the Pacific coast of the Baja California Peninsula, Mexico ¹²	0-20 ¹²	Ac, Ad ⁷⁷	A ⁷⁸ (see also ²⁴)	Yes ⁷⁹
<i>O. maya</i>	11.0-17.0 ¹²	~2,000 ¹²	M ⁵⁵	Pacific Ocean, Gulf of Mexico along the coasts of Yucatan and Campeche, Mexico ¹²	0-50 ¹²	Ac, Ad ^{15,80}	A ⁸¹ (see also ²⁴)	N/A

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Table 2. Species-specific recommendations for the accommodation and care of cephalopods species mostly utilized, or potentially utilized in Europe. For each species and a given body size we provide minimum values for water surface area and water depth, and required additional space in the case of group holding; general indication on tank shape is also provided.

Family	Species	Body length ^a (cm)	Minimum water surface area (cm ²) ^{b,c}	Minimum water surface area for each additional animal in group-holding (cm ²)	Tank shape	Minimum water depth (cm)
Nautilidae	<i>Nautilus</i> ^d	> 10	1700	400	Any shape	140
Sepiidae	<i>Sepia officinalis</i> ^e	Up to-2	100	40	Any shape	7
		2 to 6	600	200	Any shape	15
		6 to 12	1200	400	Any shape	20
		> 12	2500	1000	Any shape	25
	<i>Sepia pharaonis</i>	Up to 2	100	40	Any shape	30
		2 to 6	500	200	Any shape	60
		> 6	1400	500	Any shape	80
Sepiolidae	<i>Euprymna spp.</i> ^f	Up to 1	50	5	Any shape	5
		1 to 3	120	50	Any shape	8
		> 3	150	100	Any shape	12
Loliginidae	<i>Sepioteuthis spp.</i> ^g	Up to 10	1500	300	Cylindrical	60
		10 to 20	3500	700	Cylindrical	90
		> 20	5000	1000	Cylindrical	90
	<i>Loligo spp.</i> ^{g, h}	Up to 15	2000	400	Cylindrical	60
		15 to 25	4500	900	Cylindrical	90
		> 25	6000	1200	Cylindrical	90

Family	Species	Body length ^a (cm)	Minimum water surface area (cm ²) ^{b,c}	Minimum water surface area for each additional animal in group-holding (cm ²)	Tank shape	Minimum water depth (cm)
Octopodidae	<i>Octopus vulgaris</i> ^{h,i}	Up to 10	2000	600	Any shape	40
		10 to 20	2600	700	Any shape	50
		> 20	4000	1200	Any shape	50
	<i>Octopus maya</i> ^j	Up to 3	200	30	Any shape	20
		3 to 6	400	120	Any shape	30
		> 6	3200	900	Any shape	50

Notes to Table 2

^a Dorsal mantle length is the reference measure for body size for all species listed, except for *Nautilus* (values referred to shell diameter)

^b Juvenile stages that go through a paralarval phase shall require appropriated settings; dimensions shall be adjusted according to the scaling principle and enable animals to perform adequate movements in the water column

^c Tanks of different shapes are used to accommodate animals. Cephalopods shall have sufficient water volume for normal locomotion, taking account of their size, age, health and feeding method. Cephalopods shall be provided with an appropriate environmental enrichment, such as hiding places or bottom substrate, unless species-specific and/or behavioral traits suggest none is required

^d For *Nautilus* the body length is determined as shell diameter.

^e The minimum values shall increase in 5% if individual shelters that allow animals to find full cover are not provided.

^f Cohort up to 40 individuals. This applies to other species of sepiolids with analogous lifestyles.

^g The minimum values shall increase in 5% if non-circular tanks are used.

^h Paralarval stages shall either be excluded, or dimensions altered according to the scaling principle. Bottom dwelling.

ⁱ Animals shall be maintained isolated; in case of group housing, visual interaction between individuals is prevented; larger additional space is required for group housing octopuses if individual shelters are provided.

^j Similar parameters shall be applied to *O. bimaculoides* (no paralarval phase; similar life cycle of *O. maya*).